Fibrinolytics, enzyme inhibitors, and cancer survival

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In cancer patients, extravascular fibrin forms a provisional extracellular matrix (tumor stroma) and, in the end, this matrix is invaded by macrophages, fibroblasts, and endothelial cells to be replaced finally by granulation tissue and connective tissue.8,9 This array of events is not exclusive to tumor tissues but occurs in wound healing as well.⁴ As fibrin is generated under physiological or pathophysiological situations, in addition to serve as a structural extracellular matrix protein and to support cell adherence and cell migration,¹⁰⁻¹² it binds and thereby activates tPA (tissue-type plasminogen activator), a key serine protease of the fibrinolytic/plasminogen activation system.^{13,14} After that, tPA will interact with plasminogen and induce its conversion into the broad-band serine protease plasmin. In turn, plasmin will target fibrin and over time this will lead to the dissolution of the fibrin matrix.15,16

Interestingly, in cancer, other members of the fibrinolytic/plasminogen activation system, the urokinase-type plasminogen activator (uPA), its receptor (uPA-R, CD87), and its inhibitor PAI-1 (plasminogen activator inhibitor type-1) come into play and make use of the host plasminogen activation system to promote tumor growth and tumor cell invasiveness.¹⁷⁻²⁰ In this context it is worth mentioning that CD87 is located in the outer leaf of the plasma membrane of tumor cells, providing the anchor for binding of proteolytically active uPA. By this, focal activation of plasminogen into plasmin is achieved.²¹⁻²³ In turn, plasmin converts the zymogen pro-uPA into proteolytically active HMW-uPA. This mode of plasminogen activation is inhibited and thereby counterbalanced by PAI-1, which interacts with CD87-bound HMW-uPA to form the trimeric CD87-uPA-PAI-1 complex which is internalized by the tumor cell, stimulating tumor cell proliferation, adherence, and migration.²⁴

Although the key components in these events have been identified and biochemically characterized, the ever growing and prominent role of the fibrinolytic/plasminogen system in various disease states is putting this system into the focus of clinical investigations. In this regard it is worth mentioning that plasmin-dependent pericellular proteolysis of the extracellular matrix surrounding the tumor nests takes part in tumor cell invasion and metastasis.^{17,25,26} Especially uPA, its receptor uPAR, and the uPA inhibitor PAI-1 emerged as markers of poor prognosis in patients afflicted with solid malignant tumors.²⁷⁻³¹ In particular, elevation of uPA and/or PAI-1 protein in cancer patients indicates an elevated risk of the patient to experience early disease recurrence (metastases). Thus, shorter survival of these patients compared to cancer patients with low content of these proteolytic factors in their tumor tissue is observed.17,26,29,32

Most of the studies having investigated the clinical value of fibrinolytic factors uPA and PAI-1 in cancer patients have been conducted with breast cancer tumor specimens. Some of the key studies, encompassing thousands of patients with primary breast cancer, are depicted in Table 1. Elevation of uPA and PAI-1 in primary breast cancer tissue is not only associated with poor prognosis but is a marker for response/failure to certain cancer therapeutics, too.³³⁻³⁶ For instance, it was shown in a multicenter clinical breast cancer therapy trial that node-negative breast cancer

| Author | Year | Country | Patien | ts (N0) | Follow-up (months) | | Mode of tissue extraction | Reference |
|--------------|------|----------------------|-----------|---|-----------------------|-------------------------|------------------------------|---------------------------------|
| Jänicke | 1990 | Germany | 115 | (54) | 12.5 | uPA | Detergent extract | Fibrinolysis 4:69 |
| Schmitt | 1990 | Germany | 115 | (54) | 12.5 | uPA. tPA | Detergent extract | Blood Coag Fibrin 1:695 |
| Jänicke | 1991 | Germany | 115 | (53) | 26 | uPA, tPA, PAI-1 | Detergent extract | Sem Throm Haemost 17:303 |
| Foekens | 1992 | , The Netherlands | 671 | (273) | 48 | uPA | Cytosol fraction | Cancer Res 52:6101 |
| Jänicke | 1993 | Germany | 247 | (101) | 30 | uPA, PAI-1 | , Detergent extract | BCRT 24:195 |
| Foekens | 1994 | , The Netherlands | 657 | (273) | 48 | uPA, PAI-1 | Cytosol fraction | J Clin Oncol 12:1648 |
| Grøhndahl-H. | 1995 | Denmark | 505 | (193) | 54 | uPA, PAI-1, uPAR | , Cytosol fraction | Clin Cancer Res 1:1079 |
| Foekens | 1995 | The Netherlands | 1,012 | (460) | 71 | uPA, PAI-1, PAI-2 | , Cytosol fraction | Cancer Res 55:1423 |
| Fernö | 1996 | Sweden | 688 | (265) | 42 | uPA | , Cytosol fraction | Eur Cancer 32:793 |
| Eppenberger | 1998 | Switzerland | 305 | (159) | 37 | uPA, PAI-1 | , Cytosol fraction | Clin Oncol 16:3129 |
| Kim | 1998 | Japan | 130 | (130) | 53 | uPA, PAI-1 | , Cytosol fraction | Clin Cancer Res 4:177 |
| Кпоор | 1998 | Denmark | 429 | (178) | 61 | uPA, PAI-1 | , Detergent extract | Br Cancer 77:932 |
| Kute | 1998 | USA | 168 | (168) | 58 | uPA, PAI-1, uPAR | Cytosol fraction | BCRT 47:9 |
| Bouchet | 1999 | France | 499 | (233) | 72 uF | PA, PAI-1, PAI-2, uPA-R | , Cytosol fraction | Clin Oncol 17:3048 |
| de Witte | 1999 | The Netherlands | 865 | (434) | 100 | tPA, tPA:PAI-1 | Cytosol fraction + | Br Cancer 80:286 |
| | | | | () | | , | , Membrane fraction | 5 |
| Foekens | 2000 | The Netherlands | 2,780 | (1,405) | 88 | uPA, PAI-1, | Cytosol fraction | Cancer Res 60:636 |
| | | | _, | (,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,, | | PAI-2, uPA-R | -) | |
| Ferrero-Pous | 2000 | France | 488 | (226) | 120 | uPA | Cytosol fraction | Clin Cancer Res 6:4745 |
| de Witte | 2001 | The Netherlands | 878 | (39) | 100 | uPAR Cvtoso | / | e fractionBr Cancer 85:85 |
| Harbeck | 2001 | Germany | 276 | (130) | 109 | uPA, PAI-1 | Detergent extract | Clin Cancer Res 7:2757 |
| Jänicke | 2001 | Germany | 556 | (556) | 32 | uPA, PAI-1 | Detergent extract | INCI 93:913 |
| Konecny | 2001 | USA | 587 | (283) | 26 | uPA, PAI-1 | Detergent extract | Clin Cancer Res 7:2448 |
| Harbeck | 2002 | Germany | 761 | (269) | 60 | uPA, PAI-1 | Detergent extract | Clin Oncol 20:1000 |
| Harbeck | 2002 | Germany | 3,424 | (1736) | 83 | uPA, PAI-1 | Detergent extract | Cancer Res 62:4617 |
| Look | 2002 | European study | 8,377 | (4,676) | 79 | uPA, PAI-1 | Cytosol fraction | Natl Cancer Inst 94:116 |
| | | | -) - · · | ()) | | | Detergent extract | , |
| Lugmani | 2002 | Kuwait | 145 | (72) | 48 | uPA, tPA, PAI-1 | Cytosol fraction | Oncol Rep 9:645 |
| Bouchet | 2003 | France | 488 | (226) | 120 | uPA, PAI-1 | Cytosol fraction | Int Biol Markers 18:207 |
| Hansen | 2003 | Denmark | 228 | (101) | 144 | uPA, PAI-1 | Detergent extract | Br Cancer 88:102 |
| Look | 2003 | European study | 8,377 | (4,676) | 79 | uPA, PAI-1 | Cytosol fraction | Thromb & Haemost 90:538 |
| | | | -) - · · | ()) | | | Detergent extract | |
| Pedersen | 2003 | Denmark | 164 | (164) | 102 | uPA, PAI-1 | Detergent extract | Eur Cancer 39:899 |
| Schrohl | 2003 | Denmark | 341 | (164) | 102 | uPA, PAI-1 | Detergent extract | Mol Cell Proteomics 2:164 |
| Zemzoum | 2003 | Germany | 128 | (128) | 126 | uPA, PAI-1 | Detergent extract | Clin Oncol 21:1022 |
| Desruisseau | 2004 | France | 193 | (94) | 94 | uPA, PAI-1 | Cytosol fraction | Int Cancer 111:733 |
| Dorssers | 2004 | The Netherlands | 2,593 | (1311) | 96 | uPA, PAI-1 | Cytosol fraction | Clin Cancer Res 10:6194 |
| Manders | 2004 | The Netherlands | 576 | (576) | | JPA, PAI-1, uPA:PAI-1 | Cytosol fraction | Cancer 101:486 |
| Manders | 2004 | The Netherlands | 1,119 | (594) | | JPA, PAI-1, uPA:PAI-1 | Cytosol fraction | Cancer Res 64:659 |
| Meo | 2004 | Italy | 196 | (196) | 65 | uPA, PAI-1 | Cytosol fraction | Int Biol Markers 19:282 |
| Zhou | 2005 | Switzerland | 56 | (56) | 52 | uPA | Cytosol fraction | Int Biochem Cell Biol 37:1130 |
| Znou | 2000 | Junizerianu | 50 | (30) | 52 | ui / (| Cycosof fraction | |

Table 1. Key references demonstrating the prognostic relevance of protelytic factors of the fibrinolysis/plasminogen activation system in patients with primary breast cancer.

Cytosol fraction: Mechanical disruption of tumor tissue yields the cytosol fraction in the supernatant of the subsequent centrifugation ; Detergent extract: Mechanical disruption of tumor tissue in the presence of non-ionic detergent yields the detergent fraction in the supernatant of the subsequent centrifugation.

patients stratified by elevated uPA and/or PAI-1 do benefit from adjuvant chemotherapy, indicating that cancer patients with low uPA and/or PAI-1 should be spared the burden of chemotoxic therapy.³³ Surprisingly, the finding that both uPA and PAI-1 are indicators of poor prognosis in cancer patients is in contrast to the known, classical role of the inhibitor PAI-1 to block uPA enzymatic action. This feature may be explained by the additional, multifunctional roles of uPA and PAI-1 in cell adherence, cell motility, cell signalling, and cell proliferation.³⁷⁻⁴¹

The involvement of fibrinolytic factors in different pathophysiologies can either be direct or by (in)activation of other proteins as well as by degradation of the extracellular matrix. Understanding the pathologies associated with (anti)proteolytic action, especially the contribution of the fibrinolytic system to the disease states, and developing novel antiproteolytic therapeutic approaches is now possible owing to the availability of new biochemical and molecular biological tools and new therapeutics. Potentially, the course of the malignant disease can be altered by pharmaco-

Table 2. Diagnostic and prognostic relevance of tissue kallikreins in breast cancer assessed by measuring tissue kallikreins in tumor tissue (mRNA and/or protein).

| Kallikrei n | Method | Clinical impact and clinical applications | References | |
|----------------|----------|--|---|--|
| KLK5 Q-RT-PCR⁴ | | <u>Unfavorable prognosis</u> • overexpressed in pre/perimenopausal, node-positive patients with ER-negative tumors • independently associated with decreased DFS and OS • independent indicator of shorter DFS and OS in node-positive patients with large tumors • associated with shorter DFS in patients with low grade | Yousef, Clin Chem 48:1241; 2002 tumors | |
| KLK7 | RT-PCR | <u>Unfavorable prognosis</u> • gene expression significantly lower in breast cancer patients of low stage (I/II) and patients with positive progesterone receptors | Talieri, Thromb Haem 91:180; 2004 | |
| KLK9 | Q-RT-PCR | Favorable prognosis • overexpressed in patients with early stage disease and small tumors • independently associated with increased DFS and OS • independent indicator of prolonged DFS and OS in patients with ER and PR-negative tumors | Yousef, BCRT 78:149, 2003 | |
| hK10 | ELISA | <u>Predictive value</u> · higher hK10 levels independently associated with a poor response to tamoxifen therapy | Luo, Br J Cancer 86:1790; 2002 | |
| KLK13 | Q-RT-PCR | <u>Favorable prognosis</u> • overexpressed in older, oestrogen receptor positive pati • associated with a prolonged DFS and OS • independent indicator of longer DFS and OS in node-, ER- and PR-positive patients with low grade tumors | Chang, Br J Cancer 86:1457; 2002 ents | |
| KLK14 | Q-RT-PCR | <u>Unfavorable prognosis</u> · overexpressed in patients with advanced stage disease · independently associated with a shorter DFS and OS · independent indicator of DFS and OS in patients with positive nodal status, larger ER and PR-positive turr | Yousef, Br J Cancer 87:1287, 2002 | |
| KLK15 | Q-RT-PCR | <u>Favorable prognosis</u> • overexpressed in node-negative patients • independently associated with a longer DFS and OS • independent indicator of longer DFS and OS in | Yousef, Br J Cancer 87:1294, 2002 | |

logical intervention of the system resulting in the *in vivo* inhibition of plasmin and uPA by administering novel-types of synthetic serine protease inhibitors.⁴²⁻⁴⁹ WX-UK1, a derivative of 3-aminophenylalanine in the L-conformation with inhibitory antiproteolytic properties, is a novel small-size synthetic inhibitor directed to serine proteases such as uPA and plasmin.^{43,44} The exceptional profile of action and the safety data obtained from an *in vivo* rat breast cancer model revealed that the metastasis inhibiting effect of WX-UK1 was highly statistically significant and is not associated with side effects normally observed when applying cytotoxic cancer therapeutics, prompting the start of clinical trials with human cancer patients. Because of the different modes of action, combination

of WX-UK1 with other kinds of cancer therapeutics is feasible and could be of benefit for the cancer patient. Therefore, a series of phase I/II clinical trials involving stand-alone therapy with WX-UK1 or combination therapy of WX-UK1 with the 5-fluorouracil pro-drug capecitabine (Xeloda®) have been started in several European countries and the USA (*http://www.pancreatica.org/full_articles/f2002_10_14.html; http://www. medicalnewstoday.com/index.php?newsid=8684; http://www.lifescience.de/portal/news_detail,* 6647,,34593,detail.html).

To-date, about 180 human serine proteases, accounting for 32 % of the total proteases encoded by the human genome, have been identified 50. Of special interest in cancer research is the recent discovery

| Kallikr | ein Technique | Diagnostic value | Prognostic value | Reference |
|---------------|----------------------------------|------------------|--------------------------|--|
| KLK4 hK4 | Q-RT-PCR Immunohistochemistry | yes | unfavorable favorable | Obiezu, Clin Cancer Res 7:2380, 2001 Dong, Clin Cancer Res 7:2363, 2001 Davidson, Am J Clin Path 123:360; 2005 |
| KLK5 hK5 | RT-PCR ELISA | yes | unfavorable | Kim, Br J Cancer 84:643, 2001 Dong, Clin Cancer Res 9:1710, 2003 Yousef, BBA 1628:88, 2003- Diamandis, Tumor Biol 24:299, 2003 |
| KLK6 hK6 | Q-RT-PCR ELISA | yes | unfavorable | Tanimoto, Tumor Biol 22:11, 2001 Hoffman, Br J Cancer 87:763, 2002 Diamandis, JCO 21:1035, 2003 Diamandis, Clin Biochem 33:579, 2000 |
| KLK7 | Q-RT-PCR | yes | unfavorable | Dong Clin Cancer, Res, 9:1710, 2003 Kyriakopoulou, Clin Biochem 36:135, 2003 Tanimoto, Cancer 86:2074, 1999 |
| KLK8 hK8 | RT-PCR ELISA | yes | favorable | Magklara, Clin Cancer Res 7:806, 2001 Shigemasa, Oncol Rep 11:1153, 2004 |
| KLK9 hK10 | Q-RT-PCR ELISA | yes yes | favorable unfavorable | Yousef, Cancer Res 61:7811, 2001, Luo, Clin Cancer Res 7:2372, 2001 Luo, Cancer Res 63:807, 2003 Shvartsman, Gyn Oncol 90:44, 2003 Luo, Clinica Chimica Acta 306:111, 2001 |
| hK11 | ELISA | yes | (un)favorable | Borgono, Int J Cancer 106:605, 2003 Diamandis, Cancer Res 62:295, 2002 Diamandis, Clin Biochem 37:823, 2004 Shigemasa, Clin Cancer Res. 10:2766, 2004 |
| hK13 | ELISA | yes | favorable | Kapadia, Clinical Chem 49:77, 2003 Scorilas, J Clin Oncol 22:678, 2004 |
| KLK14 hK14 | Q-RT-PCR ELISA | yes | favorable | Yousef, Am J Clin Pathol 119:346, 2003 Borgono, Cancer Res 63:9032, 2003 Yousef, Cancer Res 61:3425, 2001 |
| KLK15 | Q-RT-PCR | yes | unfavorable | Yousef JCO 21:3119, 2003. |

Table 3. Diagnostic and prognostic relevance of tissue kallikreins in ovarian cancer assessed by measuring tissue kallikreins in tumor tissue, serum, and/or ascites (mRNA and/or protein).

of all 15 members of the human tissue kallikrein family of genes (KLK1-15), located on chromosome 19q13.4 and belonging to the S1A subfamily of serine proteases 50-53. With the exception of hK4, all tissue kallikreins have a pro-peptide ending in Lys or Arg, suggesting that these zymogens are activated by enzymes with trypsin-like activity. Intriguingly, tissue kallikreins do activate the pro-enzyme form of uPA to generate proteolytically active HMW-uPA suggesting a role of the tissue kallikreins in a proteolytic zymogen activation cascade similar to that of the blood clotting system or the complement system 51,54-56.

Tissue kallikreins, often at low levels of expression,

are found in several different organs.⁵⁰⁻⁵³ Three of the human kallikreins (KLK1-3) are called classical kallikreins because of their earlier discovery; the 12 kallikrein genes (KLK4-15) discovered during the last few years are termed *new* kallikreins. Plasma kallikrein (KLKB1), located on chromosome 4 and expressed solely in the liver, is different from the tissue kallikreins. It is involved in blood clotting, fibrinolysis, inflammatory reactions, and regulation of blood pressure. In fact, apart from plasma kallikrein, none of the 15 tissue kallikreins except the classical hK1 have appreciable kininogenase activity.

With the identification and characterization of all

members of the tissue kallikrein gene family, accumulating reports started about 4 years ago to indicate that in addition to the classical tissue kallikreins hK1-3, the *new* tissue kallikreins hK4-15 might also be related to hormonally regulated malignancies such as that of the prostate, testis, breast, and ovary 50,51. Generally, in contrast to their upregulation in ovarian cancer, tissue kallikrein genes and proteins are downregulated in cancer of the breast, prostate, and testis. A number of tissue kallikreins are also differently expressed in other types of cancer, e.g. in squamouscell carcinoma, in lung adenocarinoma, acute lymphoblastic leukaemia, and in cancer of the pancreas, head, and neck.^{50,51}

In prostate cancer, the new tissue kallikreins KLK4, 5, 10, 11, 14, 15 have been characterized at the mRNA level; hK4 and hK11 have been investigated at the protein level as well. Among the classical tissue kallikreins, hK2 and hK3 (PSA, prostate-specific antigen) are well known marker for the diagnosis, monitoring, and prognosis of prostate cancer patients. KLK5 and KLK11, determined at the mRNA level emerged as markers to predict a favorable prognosis 50,51. In testicular cancer, mRNA studies only have been conducted so far demonstrating down-regulation of KLK5, 10, 11, 13, 14 compared to normal testis tissue.⁵⁰ Regarding breast cancer (Table 2), seven out of the 12 new members of the human tissue kallikrein family are of prognostic/predictive value in breast cancer as assessed by (Q)-RT-PCR, except for hK10 expression, which was determined by ELISA. Among those tissue kallikreins

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determined by (Q)-RT-PCR, three, encompassing tissue kallikreins KLK9, 13, and 15, are markers of favorable prognosis; KLK5, KLK7, and KLK14 identify breast cancer patients with unfavorable prognosis. Tissue kallikrein hK10 is of predictive value; higher hK10 protein levels are associated with a poor response of breast cancer patients to tamoxifen therapy.

Eleven out of the 12 new members of the human tissue kallikrein family are of diagnostic/prognostic value in ovarian cancer (Table 3). Among these, 6 tissue kallikreins, encompassing tissue kallikreins hK4, 5, 6, 7, 10, 15, are markers of poor prognosis. Increase of tissue kallikreins hK8, 9, 11, 13, 14 identify ovarian cancer patients with a favorable prognosis. Tissue kallikreins hK4, hK6, and hK10 are highly expressed in serous epithelial ovarian tumors whereas higher expression of tissue kallikreins hK5, hK11, and hK13 is more frequently found in non-serous tumors. These data suggest that certain tissue kallikreins may be employed as determinants of prognosis in the subgroups of ovarian cancer patients, stratified by histiotype. Seven tissue kallikreins have been determined by ELISA (hK5, 6, 8, 10, 11, 13, 14), eight by RT-PCR (KLK4, 5, 6, 7, 8, 9, 14, 15), one by immunohistochemistry (hK4). These findings support the assumption that some of the tissue kallikreins are directly involved with cancer progression and metastasis. Tissue kallikreins therefore may represent not only novel tumor biomarkers but may also serve as promising therapeutic targets in cancer.

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