A case of *Mycobacterium goodii* infection related to an indwelling catheter placed for the treatment of chronic symptoms attributed to Lyme disease

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**Abstract**

*Mycobacterium goodii* has only rarely been reported to cause invasive disease in humans. Previously reported cases of *M. goodii* infection have included prosthetic joint infections, pacemaker pocket infections, and pneumonia. We present a case of *M. goodii* bacteremia with concomitant pulmonary septic emboli that developed in a 32-year-old woman with an indwelling central venous catheter (CVC). The CVC had been placed one year previously for intermittent treatment with intravenous, broad-spectrum antibiotics, administered by an outside physician for the treatment of symptoms attributed to chronic Lyme disease. Despite our recommendations, the patient declined follow-up in our Infectious Diseases clinic, opting to continue care under her chronic Lyme disease physician. This case clearly demonstrates the potential for serious medical complications that can arise from the inappropriate use of long-term intravenous antibiotics using a CVC to treat non-specific symptoms attributed to Lyme disease and patients should be counseled regarding these risks.

**Introduction**

*Mycobacterium goodii* is a rapidly growing non-tuberculous mycobacterium (rNTM) that was identified as a relative of *Mycobacterium smegmatis* in 1999. It is becoming recognized as an emerging cause of nosocomial infection, particularly in association with surgically implanted materials.1,2 However, only rarely have infections with *M. goodii* been associated with CVC infections. We present a case of *M. goodii* cultured from a CVC tip in the setting of persistent *M. goodii* bacteremia and pulmonary septic emboli following long-term exposure to broad-spectrum intravenous antibiotics. This is the only second reported CVC infection due to *M. goodii*, and the first case reported in a patient with extensive outpatient exposure to intravenous broad-spectrum antibiotics.

**Case Report**

A 32-year-old woman living in North Carolina but under the care of a local physician in Washington DC for symptoms attributed to chronic Lyme disease presented to George Washington University Hospital with fever, shortness of breath, and increasing confusion. Two years prior to the current presentation, the patient had been given a diagnosis of chronic Lyme disease after she developed multiple symptoms including confusion, photophobia, arthralgia/myalgia, headaches, constipation, and seizures. A computed axial tomography (CT) scan of the chest at the time showed patchy infiltrates and she underwent a bronchoscopy, which did not detect any abnormality including negative acid-fast bacilli smear and culture. Subsequent imaging showed resolution of those infiltrates. She was treated with multiple oral regimens including rifabutin, pyrimethamine, leucovorin, metronidazole and ivermectin. One year prior to her current presentation she had a subclavian CVC implanted for administration of intravenous antibiotics including meropenem, clindamycin, and tigecycline. Most recently, she had completed a three-week course of ciprofloxacin and meropenem every other day, ending several weeks prior to her presentation. After development of her current symptoms, she sought treatment in an emergency department in North Carolina, where she was diagnosed with pneumonia and treated with azithromycin without improvement. She then traveled to Washington, DC for evaluation by her local physician, and subsequently presented to our institution.

In the emergency department the patient was noted to be febrile to 39.5°C, blood pressure 125/73 mmHg, heart rate 85 beats/minute. Physical exam revealed fatigue appearance with decreased breath sounds bilaterally, and a CVC with port access in the left chest wall without evidence of infection. Laboratory studies revealed leukocytosis (white blood count 11,370 cells/mL, 85% neutrophils). Contrast CT chest showed bilaterally symmetric opacification with bronchovascular distribution and innumerable pulmonary nodules (Figure 1). Multifocal pneumonia due to septic emboli was suspected, and vancomycin plus piperacillin-tazobactam were started. A trans-thoracic echocardiogram (TTE) found no evidence of valvular vegetations. No other foci of infection were apparent on physical examination. After 72 hours anaerobic blood culture bottles (BACTEC Plus Aerobic/F, Becton Dickinson, Sparks, Maryland) revealed growth of beaded Gram-positive, acid-fast rods suggestive of mycobacterial species.

After lengthy discussion with the patient and her family, the CVC was removed. Culture of initial blood cultures and the CVC tip revealed growth of *M. goodii*. The patient’s antibiotics were...
empirically changed to azithromycin 500 mg intravenously daily, ciprofloxacin 400 mg intravenously every 12 hours, and meropenem 1 g intravenously every 8 hours. The patient clinically improved on this regimen. The patient returned to North Carolina before the identification and sensitivities of the mycobacterial species was available. Therefore, she was empirically switched to an oral regimen of trimethoprim/sulfamethoxazole 800 mg/160 mg twice daily, azithromycin 250 mg once daily, and ciprofloxacin 500 mg twice daily for two-weeks from time of line removal. The patient was counseled extensively to avoid future placement of a CVC in the absence of a clear medical indication. Blood cultures drawn after CVC removal were negative at the time of her discharge but subsequent to discharge also grew *M. goodii*. The patient cancelled her follow-up appointment in our Infectious Diseases clinic and she was lost to follow-up.

The mycobacterial isolate was sent to Quest Diagnostics Labs (San Juan Capistrano, California) for the beta subunit

![Computed axial tomography of the chest performed with intravenous contrast showing bilateral nodular opacifications in a bronchovascular distribution.](image-url)

**Figure 1.**

**Table 1.** Cases of rapid-growing non-tuberculosis mycobacteria (rNTM) identified in blood cultures at our institution, 2012-2018.

<table>
<thead>
<tr>
<th>Case</th>
<th>Month/Year</th>
<th>Organism</th>
<th>Central line</th>
<th>Line removed</th>
<th>Culture clearance</th>
<th>Antibiotics</th>
<th>Outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>59-year-old man with ESRD</td>
<td>09/2018</td>
<td><em>M. chelonae</em></td>
<td>CVC, tunneled</td>
<td>Y</td>
<td>Y</td>
<td>ciprofloxacin and metronidazole for 7 days</td>
<td>Infection resolved</td>
</tr>
<tr>
<td>32-year-old woman with “Chronic Lyme disease”</td>
<td>03/2018</td>
<td><em>M. goodii</em></td>
<td>CVC, port</td>
<td>Y</td>
<td>N</td>
<td>See text</td>
<td>Declined follow-up</td>
</tr>
<tr>
<td>70-year-old man with Parkinson’s Disease</td>
<td>02/2018</td>
<td><em>M. mucogenicum</em></td>
<td>PICC</td>
<td>Y</td>
<td>Y</td>
<td>piperacillin/tazobactam for 2 weeks</td>
<td>Infection resolved</td>
</tr>
<tr>
<td>28-year-old man with Crohn’s disease</td>
<td>12/2017</td>
<td><em>M. mucogenicum</em></td>
<td>No midline</td>
<td>Y</td>
<td>Y</td>
<td>linezolid for 2 weeks</td>
<td>Infection resolved</td>
</tr>
<tr>
<td>27-year-old woman with ESRD</td>
<td>10/2016</td>
<td><em>M. fortuitum</em></td>
<td>CVC, tunneled</td>
<td>Y</td>
<td>Y</td>
<td>ciprofloxacin and clarithromycin, followed by ciprofloxacin and linezolid for 3 months</td>
<td>Infection resolved</td>
</tr>
<tr>
<td>70-year-old woman with Crohn’s disease and recurrent urinary tract infections</td>
<td>08/2016 and 04/2018</td>
<td><em>M. fortuitum</em> complex</td>
<td>CVC, PICC</td>
<td>Y</td>
<td>Y*</td>
<td>ciprofloxacin for 2 weeks</td>
<td>Cleared initially but recurred 1.5 years later with eventual clearance after second PICC line removal</td>
</tr>
<tr>
<td>31-year-old woman with renal transplant</td>
<td>02/2016</td>
<td><em>M. mucogenicum</em> group</td>
<td>CVC, port</td>
<td>Y</td>
<td>Y</td>
<td>meropenem, unclear duration</td>
<td>Infection resolved</td>
</tr>
<tr>
<td>43-year-old woman with native-valve endocarditis</td>
<td>12/2015</td>
<td><em>M. mucogenicum</em> group</td>
<td>PICC</td>
<td>Y</td>
<td>Y</td>
<td>cefepime for 6 weeks</td>
<td>Infection resolved</td>
</tr>
<tr>
<td>52-year-old man with HIV and recurrent urinary tract infections</td>
<td>09/2013</td>
<td><em>M. mucogenicum</em> group</td>
<td>PICC</td>
<td>Y</td>
<td>Y</td>
<td>piperacillin/tazobactam and daptomycin for 4 weeks</td>
<td>Infection resolved</td>
</tr>
<tr>
<td>27-year-old woman with sickle cell anemia</td>
<td>11/2012</td>
<td><em>M. mucogenicum</em> group</td>
<td>CVC, internal jugular</td>
<td>Y</td>
<td>Y</td>
<td>ciprofloxacin and linezolid for 2 weeks</td>
<td>Infection resolved</td>
</tr>
</tbody>
</table>

ESRD = end-stage renal disease, CVC = central venous catheter, PICC = peripherally inserted central catheter, Midline = peripherally inserted catheter, Y = blood cultures were drawn after line was removed and were immediately negative, N = blood cultures were not negative after line removal. *Two instances (see Outcome).
Discussion

rNTMs are hydrophobic, ubiquitous organisms found in water and soil. They have been observed to form biofilms in non-stagnant water piping, survive a variety of environmental extremes (temperature, pH, scarcity of nutrients), and are regarded as resistant to disinfectants such as chlorine and phenol/glutaraldehyde based substances. \(^3\) *M. goodii* has been associated with soft tissue infection, osteomyelitis, bursitis, pneumonia, and achalasia-associated pulmonary disease.\(^4,5\) Cases of *M. goodii* infections have also been reported to complicate a variety of surgical procedures and devices, including: hardware,\(^6\) pacemaker devices,\(^7\) mitral valve annuloplasty,\(^8\) prosthetic heart valves,\(^9\) deep brain stimulators,\(^10\) intra-ophthalmic devices,\(^11\) intra-abdominal mesh,\(^12\) ventriculoperitoneal shunts,\(^13\) breast implants,\(^14\) and intravenous catheters.\(^5\) In the majority of these cases involving prosthetic material, clearance of the infection was only achieved by removal of foreign material and administration of antibiotic therapy. In only two cases was antibiotic therapy alone successful despite retention of hardware: one involving a deep brain stimulator,\(^15\) and another a pacemaker pocket infection.\(^5\)

A retrospective review of blood cultures positive for rNTM at our institution over the last six years revealed ten episodes of rNTM bacteremia in nine patients (in addition to the case presented above) summarized in Table 1. These included three cases of *M. fortuitum* complex, one case of *M. chelonae*, and six cases of *M. mucogenicum*. In all cases, a CVC was implicated, and subsequently removed. All cases were treated antibiotics, although the regimen and duration of treatment varied. With the exception of our case, clearance of the bacteremia followed CVC removal. All of the above isolates grew in standard BACTEC blood culture bottles, with subsequent incubation on Löwenstein Jensen media at 36°C with 5% CO\(_2\). All specimens were sent to Quest Diagnostics for identification after in-house DNA probes were negative for *M. tuberculosis* complex, *M. avium* complex, and *M. kansasii*.

Catheter-associated bloodstream infections caused by rNTM are seen most frequently in patients with malignancies that require CVC for chemotherapy. These infections typically have greater than 90% clearance rate with catheter removal even in the absence of antibiotic therapy.\(^13,14\) Unlike the majority of cases of rNTM catheter-associated infections, our patient did not clear her blood cultures immediately after catheter removal. This may have been due to more persistent foci of infection from the septic emboli in her lungs, or possibly due to an infected thrombus at the site of her former long-term CVC. Upper-extremity thrombosis has been described as a risk with the use of catheters similar to that used in our patient, with one study reporting 3 times higher risk for such outcomes.\(^15\) While mycobacterial native valve endocarditis is exceedingly rare,\(^16\) and a good-quality TTE was negative in our case, the suspicion of secondary foci of infection warranted close follow-up in this patient, and possible extension of treatment duration for longer than two weeks. Unfortunately, the patient did not return for outpatient follow-up or additional imaging.

Macrolide resistance is a common concern when treating rNTM,\(^17\) and was documented in our isolate; the patient’s history of receiving azithromycin monotherapy prior to her presentation to our hospital may have contributed to this. The empirical multidrug regimen that we started, including the intravenous ciprofloxacin, azithromycin, and sulfamethoxazole/trimethoprim as an outpatient, was supported by the susceptibility data and should have been adequate therapy. Due to persistent symptoms, patients with post-Lyme disease syndrome (PLDS) may seek alternative therapies that have limited supporting evidence and can be associated with significant morbidity. Long-term antibiotic therapy has not been shown to have any benefit for PLDS,\(^18\) but is a clear risk factor for catheter infections and selection of resistant organisms. The use of antibiotics for the treatment of PLDS, especially when given intravenously, has been associated with increased hospitalization, emergency department visits, and infection.\(^19\) To date, four cases of serious bloodstream infections associated with catheters placed specifically for long-term antibiotic treatment of symptoms attributed to Lyme disease have been reported, with one case resulting in death.\(^19\) Our patient represents the first case of PLDS treatment complicated by a mycobacterial infection. As illustrated, the identification and treatment of rNTM organisms can pose particular challenges including multi-drug resistance (especially in patients already exposed to multiple antibiotics), reliance on specialty laboratory methods, and longer times to definitive identification and sensitivity testing. Ultimately, there was a significant trust gap between the patient and our infectious disease physicians, and the patient stated her intention to keep following with her “Lyme-literate” provider and was lost to follow-up.

Conclusions

Mycobacterial infections, including *M. goodii*, should be considered as potential causes of CVC infections facilitating optimal diagnostic testing and use of appropriate antibiotic regimens. Our patient’s CVC and long-term use of antibiotics selected for an unusual biofilm-producing pathogen: *M. goodii*. In contrast to published literature as well as to prior experience with rNTM bloodstream infections at our institution, blood cultures remained positive even after CVC removal and initiation of appropriate antimicrobial therapy, suggesting the persistence of a persistent nidus of infection. The patient was counseled on the need for close outpatient follow-up with an infectious disease specialist and avoidance of future long-term line placement without clear medical necessity.

References


